

スギ花粉に含まれる内生ジベレリン

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Endogenous gibberellins in the pollen of *Cryptomeria japonica* D. DON

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HONMA, T., YAMAGUCHI, I., NAKAJIMA, M., MUROFUSHI, N. and MIGITA, K.: **Endogenous gibberellins in the pollen of *Cryptomeria japonica* D. DON.** *J. Jpn. For. Soc.* 77: 358~365, 1995 Shoot elongation and flower bud formation of sugi (*Cryptomeria japonica* D. DON) are promoted by exogenous gibberellins (GAs). This suggests that these physiological events are regulated by endogenous GAs, too. Little is known about the endogenous GAs of conifers. GAs are characterized from only four species of Pinaceae. Although the occurrence of GA-like substances in *C. japonica* was shown by bioassay using dwarf rices, their identification has not been reported. As the first step to study the relationship between endogenous GAs and the physiological and morphological changes in the growth of *C. japonica*, we analyzed endogenous GAs in the pollen of this plant. The occurrence of GA₁ and/or GA₃, GA₄, GA₉, GA₁₂, and GA₁₅ was suggested by combination of bioassays using dwarf rice (*Oryza sativa* L. cv. Tan-ginbozu) and enzyme-linked immunosorbent assay (ELISA) after purification by high performance liquid chromatography (HPLC). Eventually, GA₉, GA₁₂, and GA₁₅ were conclusively identified by gas chromatography-mass spectrometry (GC/MS). However, the identification of GA₁ and/or GA₃ and GA₄ by either GC/MS or gas chromatography-selected ion monitoring (GC/SIM) was unsuccessful, because these amounts were too small for the detection. This result shows that GAs in the early-non-hydroxylation biosynthetic pathway are predominant in the pollen of *C. japonica*.

本間 環・山口五十麿・中嶋正敏・室伏 旭・右田一雄：スギ花粉に含まれる内生ジベレリン 日林誌 77: 358~365, 1995 スギ (*Cryptomeria japonica* D. DON) の伸長成長および花芽形成は、他の針葉樹と同様にジベレリン (GAs) の投与によって促進される。このことは、これらの生理現象が内生 GAs によって調節されている可能性をも示している。しかし、針葉樹の内生 GAs に関する知見は少ない。これまでに、針葉樹の内生 GAs の報告はマツ科の 4 種に限られている。一方、スギのジベレリン様物質は矮性イネ「短銀坊主」によるバイオアッセイにより確認されているものの、その同定は未だ報告されていない。本研究では、内生 GAs とスギの生理学および形態学的変化との関連を解明する研究の第一段階として、花粉に含まれる内生 GAs の分析を行った。その結果、高速液体クロマトグラフィーにより精製した試料を「短銀坊主」によるバイオアッセイおよびエンザイム免疫アッセイ (ELISAs) を用いて分析し、GA₁ および/または GA₃, GA₄, GA₉, GA₁₂, GA₁₅ の存在を推測した。それらのうち GA₉, GA₁₂ および GA₁₅ をガスクロマトグラフィー質量分析 (GC/MS) により同定した。しかしながら、ELISA によりその存在が示唆された GA₁ および/または GA₃ と GA₄ については、含有量が少ないために GC/MS あるいは GC/SIM のいずれにおいても同定することはできなかった。これらの結果から、スギの花粉には early-non-hydroxylation pathway が主要な生合成経路として機能している可能性が示された。

I. Introduction

The physiological effects of exogenously applied gibberellins (GAs) on conifers have been studied by many research groups (PHARIS and KING, 1985). Exogenously applied GAs promote shoot elongation and flower bud formation in many species of conifers. KATO *et al.* (1958) reported that exogenously applied GAs

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(gibberellin powder) promoted flower bud formation of sugi (*Cryptomeria japonica* D. DON), a species of Taxodiaceae, and HASHIZUME (1959) reported the promotion of shoot elongation of the same species by exogenously applied GAs. These observations suggest that endogenous GAs are involved in these physiological events in nature. The occurrence of GA-like substances in 31 conifer species were reported by DUNBERG and ODÉN (1983). However, endogenous GAs were identified from only four species of Pinaceae, and little is known about the presence of endogenous GAs of other species. We are interested in the relationship between these physiological events and the dynamics of endogenous GAs of *C. japonica*. The occurrence of GA-like substances in this plant was reported by HASHIZUME (1961) and OGIYAMA and PHARIS (1980) based on the biological activities in dwarf rice bioassays, but none of them was identified. Thus, we tried to identify endogenous GAs of this species as many as possible for the future study on the relationship between its physiological events and the dynamics of endogenous GAs. The pollen was selected as the first material for the survey of GAs, because the concentration of GAs was expected to be high in the pollen of *C. japonica* as in *Pinus attenuata* LEMM. (KAMIENSKA *et al.*, 1976a). In this paper, the GAs in the pollen of *C. japonica* were carefully surveyed by combination of dwarf rice bioassay, enzyme-linked immunosorbent assay (ELISA) and, gas chromatography-mass spectrometry (GC/MS).

II. Materials and Methods

1. Plant materials

Branches with male flowers collected just before blooming at the Chiba Experimental Forest of the University of Tokyo in February 1993, were kept in water pails at 18°C for 14 days until full bloom. The pollen was collected from the fully open anthers of male flowers, and stored at -85°C until processed for analysis.

2. Processing of the pollen for analysis

The purification procedure of the pollen is shown in Fig. 1.

1) Extraction and solvent fractionation—The pollen (700 g fresh weight, fr wt) was soaked and homogenized in 80% acetone (v/v) and extracted with the same solvent. After, filtration, the extracts were concentrated *in vacuo*. The aqueous concentrate was subjected to a solvent fractionation, and an acidic ethyl acetate (AE) fraction was obtained (YAMAGUCHI *et al.*, 1990).

2) High performance liquid chromatography (HPLC)—The dried AE fraction was dissolved in a small volume of methanol (MeOH) and loaded on a Bond Elut DEA cartridge (18 g 40 μ m, Analytichem International Co., Ltd., USA.), which was eluted with MeOH (20 ml) and 0.5% acetic acid (AcOH) in MeOH (30 ml), successively. The eluates of the acidic MeOH were concentrated and subjected to HPLC (L-6000 pump and L-6200 intelligent pump, Hitachi Ltd., Tokyo) of an Octadecylsilan (ODS) column (15 cm \times 6 mm inside diameter, SCC-ODS-H2151, Senshu Scientific Co., Tokyo) using the following gradient elution between solvents A (acetonitrile (MeCN) : H₂O, 2 : 8, containing 0.5% AcOH) and solvent B (MeCN : H₂O, 8 : 2, containing 0.5% AcOH) ; 0~5 min : isocratic solvent A, 5~35 min : linear gradient from 100% solvent A to 100% solvent B, 35~45 min : isocratic solvent B. The flow rate was 1.0 ml/min, and the eluates were collected in 1 min fractions.

3) Dwarf rice bioassay—Biological activity in each fraction of ODS-HPLC was assayed by the micro-drop method (MURAKAMI, 1968) using the dwarf rice (*Oryza sativa* L. cv. Tan-ginbozu). The concentrate of each fraction was dissolved in 50% aqueous acetone (v/v), and an aliquot (20 g fr wt equivalent (equiv.)) was applied to each plant.

4) Enzyme-linked immunosorbent assay (ELISA)—ELISA was conducted in the same manner as reported by YAMAGUCHI *et al.* (1990). An anti-GA₁-Me- or an anti-GA₂₀-Me-antiserum and GA-labeled alkaline phosphatase as a tracer (YAMAGUCHI *et al.*, 1987, 1990) were used for each assay. The cross-reactivities of each GA-Me against the anti-GA₁-Me- and the anti-GA₂₀-Me-antiserum are summarized in Fig. 2. A portion of each fraction from ODS-HPLC was concentrated, methylated with diazomethane (CH₂N₂), dried, and re-dissolved in 5% MeOH in 50 mM tris buffer (pH 7.6). An aliquot (3 g fr wt equiv.) of each methylated fraction was used for the assays. The absorbance at 410 nm of nitrophenol was measured by a microplate

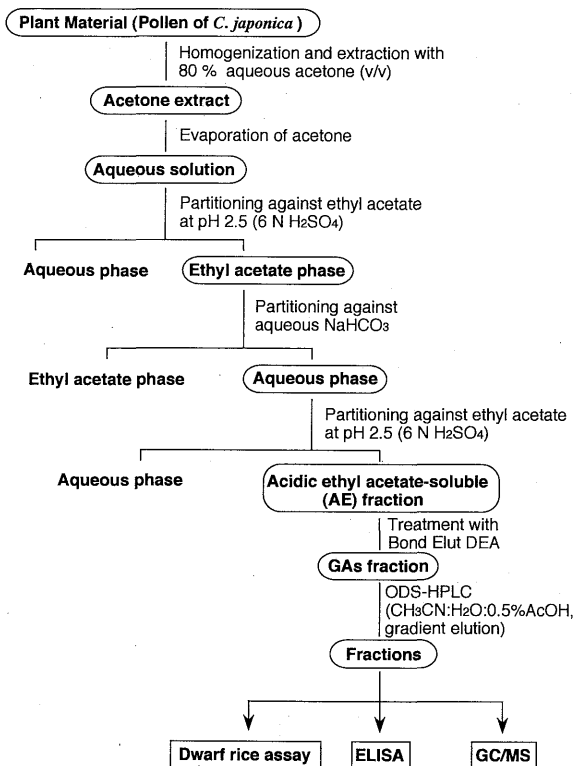


Fig. 1. Procedure for the extraction, purification and identification of GAs in the pollen of *C. japonica*

reader (MR 5000, Dynatech, USA.).

5) Gas chromatography-mass spectrometry (GC/MS)—A Hitachi M-80B mass spectrometer equipped with a Hewlett Packard 5980 Agas chromatograph and a capillary column Ultra # 1 (25 m × 0.2 mm *i. d.*, Hewlett Packard, USA.) was used under the following conditions; injection temperature: 250°C, column temperature: 60°C (1.5 min), linear gradient up to 200°C at 60°C/min, to 280°C at 5°C/min, and kept at 280°C (10 min). Ionization voltage was 70 eV. The remainder of each fraction from ODS-HPLC were subjected to GC/MS and/or GC/SIM after methylation with CH₂N₂ and trimethylsilylation with *N*-methyl-*N*-trimethylsilyltrifluoroacetamide. In GC/SIM, characteristic ions, *m/z*, (mass to charge ratio peak) 506 (M⁺, molecular ion peak), 491, and 448 for GA₁, *m/z* 504 (M⁺), 489, and 447 for GA₃, and *m/z* 418 (M⁺), 390, and 386 for GA₄, were monitored. Hydrocarbons C₁₈ to C₃₀ with even numbers of carbon atoms, were used for the calculation of the modified Kovats retention indices for the identification of GAs.

III. Results

Figure 3 shows a histogram of biological activities on Tan-ginbozu of the fractions from ODS-HPLC. Three peaks were observed in the fractions of, retention times (*t_R*) 25~26 min, 29~30 min, and 31~33 min, GA₅, GA₂₀, and GA₄₄ were put up for the candidates of the biologically active substances at *t_R* 25~26 min, GA₉ and GA₁₅ to those at *t_R* 29~30 min and GA₁₂ to that at *t_R* 31~33 min.

A histogram of immunoreactivities against the anti-GA₁-Me-antiserum of the fractions from ODS-HPLC is shown in Fig. 4. Two clear immunoreactive peaks were observed at *t_R* 4~5 and 10~11 min with some small peaks scattered over the fractions of *t_R* 15~19, 23~27, and 32~35 min. Judging from the *t_R* and the specificity of the antiserum, GA₁ and GA₃ could be put up as candidates of the active principles at *t_R* 10~11

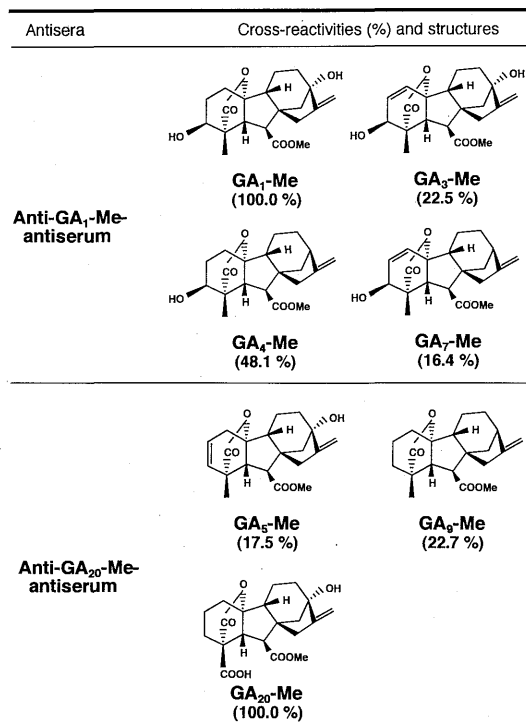


Fig. 2. Structures of GA methyl esters showing high cross-reactivity against an anti-GA₁-Me- and an anti-GA₂₀-Me-antiserum

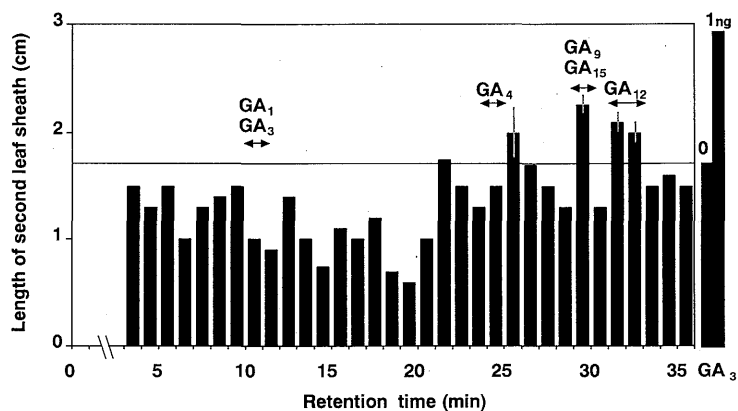


Fig. 3. Histogram of growth promoting activities on dwarf rice (Tan-ginbozu) of the fractions from ODS-HPLC of the AE fraction of the pollen (*C. japonica*).

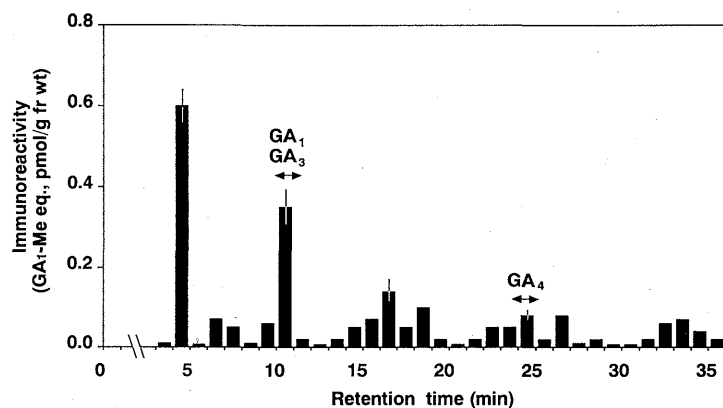


Fig. 4. Histogram of immunoreactivities against anti-GA₁-Me- anti-serum of the fractions from ODS-HPLC of the AE fraction of the pollen (*C. japonica*).

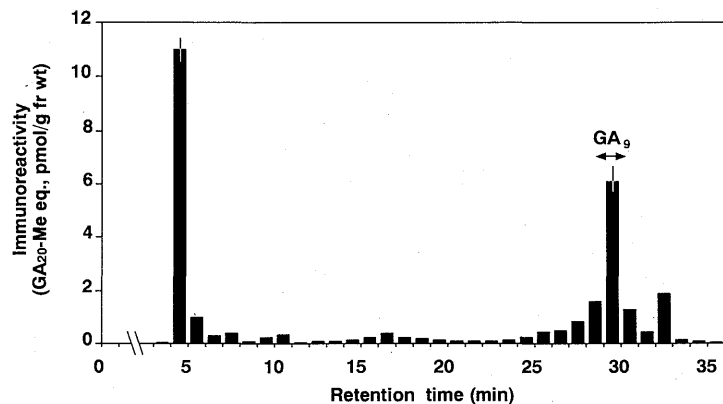


Fig. 5. Histogram of immunoreactivities against anti-GA₂₀-Me- anti-serum of the fractions from ODS-HPLC of the AE fraction of the pollen (*C. japonica*).

Table 1. Retention times (ODS-HPLC and GC), modified Kovats retention indices (KRI), and characteristic ions of GAs identified from the pollen of *C. japonica*

GAs		Retention time (min)		KRI	Characteristic ions, <i>m/z</i> (relative intensity)
		HPLC	GC		
GA ₉	Sample	29~30	10.37	2349	300[M ⁺]. (8), 298(69), 270(100)
	Authentic		10.45	2353	330[M ⁺]. (10), 298(83), 270(100), 243(60)
GA ₁₅	Sample	29~30	13.95	2665	344[M ⁺]. (18), 312(23), 284(68), 239(100)
	Authentic		13.98	2667	344[M ⁺]. (19), 312(23), 284(68), 239(100)
GA ₁₂	Sample	32~33	10.65	2404	360[M ⁺]. (0), 328(20), 300(100), 285(25), 240(42)
	Authentic		10.70	2410	360[M ⁺]. (0), 328(20), 300(100), 285(25), 240(39)

min, but none for that at t_R 4~5 min. At the elution zone of GA₄ (t_R 23~25 min), only a small immunoreactivity was observed. The other histogram of immunoreactivities against anti-GA₂₀-Me-antiserum is shown in Fig. 5. Prominent immunoreactive peaks were observed at t_R 4~5 min and 29~30 min with a medium one at t_R 32~33 min. Judging from the t_R and the specificity of the anti-GA₂₀-Me-antiserum, GA₉ was put up as the candidate for the immunoreactive substance at t_R 29~30 min. There were no candidates for the activity at t_R 4~5 min, because the t_R is too small for known GAs with high cross-reactivity to anti-GA₂₀-Me-antiserum. The possibility of GA₅ and GA₂₀ as the candidates of biological activity on dwarf rice at t_R 25~26 min was excluded because no immunoreactivity was detected by ELISA using anti-GA₂₀-Me-antiserum.

To identify GAs definitely, the fractions which showed either biological activity in dwarf rice assay or immunoreactivity against anti-GA₁-Me- or anti-GA₂₀-Me-antiserum were subjected to full scan GC/MS or GC/SIM. GA₉, GA₁₂, and GA₁₅ were identified based on the modified KOVATS' retention index and full scan mass spectra. The details are summarized in Table 1. The fractions of t_R 10~11 min and t_R 24~25 min, which corresponded to the elution zones of GA₁ and/or GA₃ and GA₄ and showed immunoreactivities against anti-GA₁-Me-antiserum, were examined by GC/SIM. However, none of the expected GAs was identified because of their small quantities.

IV. Discussion

The first identification of endogenous GAs from conifers by GC/MS was reported by KAMIENSKA *et al.* (1976a). They identified GA₃, GA₄, and GA₇ from the pollen of *Pinus attenuata*. GA₉ was identified from shoots of *Picea abies* L. KARST. (ODÉN *et al.*, 1982) and GA₁, GA₃, and GA₉ were identified from shoots of the same species (ODÉN *et al.*, 1987). GA₁, GA₄, and GA₉ were identified from shoots of *Picea sitchensis* BONG. CARR. during the period of cone-bud differentiation (MORIZ *et al.*, 1989a). GA₁, GA₃, GA₄, and GA₉ were identified from shoots of *Picea sitchensis* during the period of flower-bud differentiation and shoot elongation (MORIZ *et al.*, 1990a, 1990b). GA₁, GA₃, GA₄, GA₇ and GA₉ were identified from shoots of *Pseudotsuga menziesii* MIRB. FRANCO. (DOUMAS *et al.*, 1992). Glucosyl esters of GA₉ and *iso*-GA₉ were identified from needle of *Picea sitchensis* (LORENZI *et al.*, 1976, 1977). A conjugate of GA₉ was identified from shoots of *Picea sitchensis* (MORIZ *et al.*, 1990a, 1990b). Glucosyl ester of GA₉ was identified from shoots of *Pseudotsuga menziesii* (DOUMAS *et al.*, 1992). These reports show that both the early-13-hydroxylation pathway and the early-non-hydroxylation pathway of GA-biosynthesis may operate in these conifers. However, no evidence of the organ specific occurrence of endogenous GAs has not been reported in conifers. In the survey of endogenous GAs in the pollen of *C. japonica*, we identified GA₉, GA₁₂, and GA₁₅ all of which are members in the early-non-hydroxylation biosynthetic pathway (Fig. 6). The occurrence of GA₁ and/or GA₃ was also suggested by ELISA (Fig. 4) after ODS-HPLC, but neither of these GAs were detected by GC/MS or GC/SIM. However, judging from the earlier reports describing the distribution of GA₁ and/or GA₃ in all conifers so far examined, the possibility of the occurrence of GA₁ and/or GA₃ at low levels in the pollen of *C. japonica* remains. Judging from the results of ELISA (Fig. 4), the content of GA₄, which is expected to be an active GA in the early-non-hydroxylation pathway must be less than a half of GA₁ and/or GA₃, if

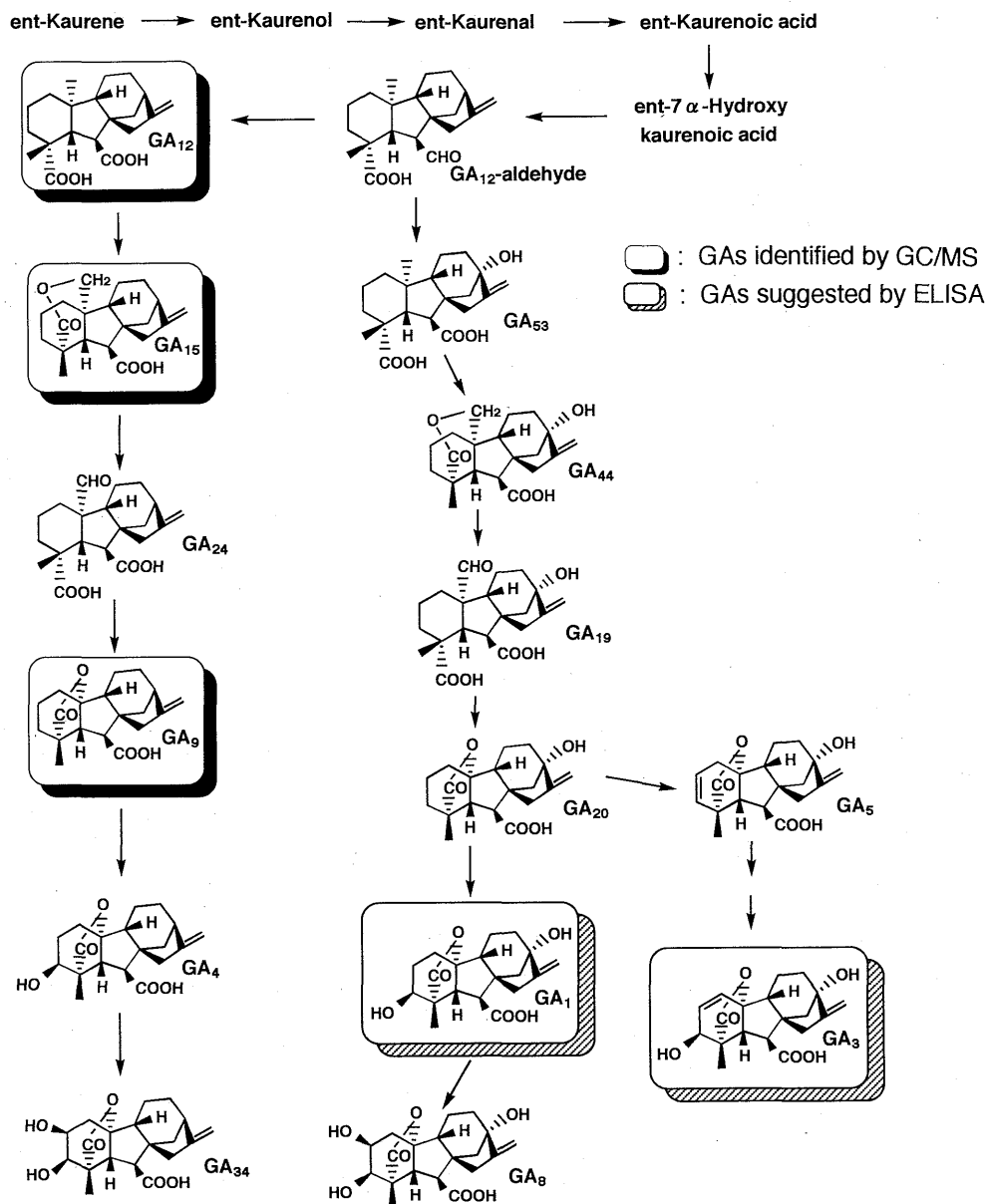


Fig. 6. Early-13-hydroxylation and early non-hydroxylation pathways of GA-biosynthesis

any. Nevertheless, GAs in the early-non-hydroxylation biosynthetic pathway are predominant in the pollen. This suggests that either the early-non-hydroxylation pathway is the major biosynthetic pathway in *C. japonica*, as in cucumber (GRAEBE, 1987) or that the GAs in this pathway distribute organ specificity in the pollen as in anthers of rice (KOBAYASHI *et al.*, 1984).

An immunoreactivity was detected in the fraction of t_R 4~5 min from ODS-HPLC by both ELISAs using anti-GA₁-Me-antibody and anti-GA₂₀-Me-antibody. Although it is impossible to deduce the immunoreactive principle in this fraction from available data, unknown GA conjugates, which show cross-reactivities to both antibodies, may be contained in the fraction as in the case of anthers of rice (HASEGAWA *et al.*, 1994).

In the present study, we could not identify 3 β -hydroxylated C₁₉-GAs, which are expected to be active GAs, from the pollen of *C. japonica*, although the occurrence of GA₁ and/or GA₃ was strongly suggested by ELISA. So far, GA₂₀ which is a direct precursor of GA₁ in the early-13-hydroxylation biosynthetic pathway has not been identified in many conifers, inspite of GA₁ and/or GA₃ having been identified from several species. This may suggest that a biosynthetic pathway of late 13-hydroxylation operates as a characteristic of conifers. Exogenously applied GA₄ was metabolized into GA₁ and GA₃₄ in germinating pollen of *Pinus attenuata* (KAMIENSKA *et al.*, 1976b). Exogenous applied GA₉ was metabolized into GA₄ and GA₁, and GA₄ into GA₁ in *Picea sitchensis* (MORIZ *et al.*, 1989b). These observations suggested the possibility that GA₁ is formed by the late 13-hydroxylation of C₁₉-GAs in Pinaceas. The same pathway may operate in *C. japonica*, too. As it is important to clarify the distribution of GAs in organs and their biosynthesis and translocation for the study of the role of GAs in the expression of physiological phenomena of *C. japonica*, we now are extensively surveying 3 β -hydroxylated GAs and 13-hydroxylated GAs in this plant.

Literature cited

- DUNBREG, A. and ODÉN, P. C. (1983) Gibberellins and conifers. *In* The biochemistry and physiology of gibberellins (CROZIER, A., ed.). Vol. 2, 221~295. Praeger, New York.
- DOUMAS, P., IMBULT, N., MORIZ, T. and ODÉN, P. C. (1992) Detection and identification of gibberellins in Douglas fir (*Pseudotsuga menziesii*) shoots. *Physiol. Plant.* **85**: 485~494.
- GRAEBE, J. E. (1987) Gibberellins biosynthesis and control. *Ann. Rev. Plant Physiol.* **38**: 419~465.
- HASEGAWA, M., NAKAJIMA, M., TAKEDA, K., YAMAGUCHI, I. and MUROFUSHI, N. (1994) A novel gibberellin glucoside, 16 α , 17-dihydroxy-16, 17-dihydrogibberellin A₄-17-O- β -D-glucopyranoside, from rice anthers. *Phytochemistry* **37**: 629~634.
- HASHIZUME, H. (1959) The effect of gibberellin upon flower formation in *Cryptomeria japonica*. *J. Jpn. For. Soc.* **41**: 375~381. (in Japanese)
- HASHIZUME, H. (1961) The effect of gibberellin on flower-bud formation in *Cryptomeria japonica*. III. Changes of endogenous growth substance, carbohydrates and nitrogen in new shoots in relation to flower induction by gibberellin. *J. Jpn. For. Soc.* **43**: 120~126. (in Japanese)
- KAMIENSKA, A., DURLEY, R. C. and PHARIS, R. P. (1976a) Isolation of gibberellins A₃, A₄ and A₇ from *Pinus attenuata* pollen. *Phytochemistry* **15**: 421~424.
- KAMIENSKA, A., DURLEY, R. C. and PHARIS, R. P. (1976b) Endogenous gibberellins of pine pollen. III. Conversion of 1, 2-[³H]GA₄ to gibberellins A₁ and A₃₄ in germinating pollen of *Pinus attenuata* LEMM. *Plant Physiol.* **58**: 68~70.
- KATO, Y., MIYAKE, I. and ISHIKAWA, H. (1958) Stimulation of gibberellin on differentiation of "Sugi." *J. Jpn. For. Soc.* **40**: 35~36. (in Japanese)
- KOBAYASHI, M., YAMAGUCHI, I., MUROFUSHI, N., OTA, Y. and TAKAHASHI, N. (1984) Endogenous gibberellins in immature seeds and flowering ears of rice. *Agric. Biol. Chem.* **48**: 2725~2729.
- LORENZI, R., HORGAN, R. and HEALD, J. K. (1976) Gibberellin A₉ glucosyl ester in needles of *Picea sitchensis*. *Phytochemistry* **15**: 789~790.
- LORENZI, R., SANDERS, P. F., HEALD, J. K. and HORGAN, R. (1977) A novel gibberellin from needles of *Picea sitchensis*. *Plant Sci. Lett.* **8**: 179~182.
- MORIZ, T., PHILIPSON, J. J. and ODÉN, P. C. (1989a) Detection and identification of gibberellins in sitka spruce (*Picea sitchensis*) of different ages and coning ability by bioassay, radioimmunoassay and gas chromatography-mass spectrometry. *Physiol. Plant.* **75**: 325~332.
- MORIZ, T., PHILIPSON, J. J. and ODÉN, P. C. (1989b) Metabolism of tritiated and deuterated gibberellins A₁, A₄ and A₉ in Sitka spruce (*Picea sitchensis*) shoots during the period of cone-bud differentiation. *Physiol. Plant.* **77**: 39~45.
- MORIZ, T., PHILIPSON, J. J. and ODÉN, P. C. (1990a) Quantification of gibberellins A₁, A₃, A₄, A₉ and A₉-conjugate in good and poor-flowering clones of Sitka spruce (*Picea sitchensis*) during the period of flower-bud differentiation. *Planta* **181**: 538~542.
- MORIZ, T., PHILIPSON, J. J. and ODÉN, P. C. (1990b) Quantification of Gibberellins A₁, A₃, A₄, A₉ and a Putative A₉-conjugate in grafts of Sitka spruce (*Picea sitchensis*) during the period of shoot elongation. *Plant Physiol.* **93**: 1476~1481.
- MURAKAMI, Y. (1968) The microdrop method, a new rice seedling test for gibberellins and its use for testing extracts of rice and morning glory. *Bot. Mag.* **81**: 33~34.
- ODÉN, P. C., ANDERSSON, B. and GREF, R. (1982) Identification of gibberellins A₉ in extracts of Norway spruce (*Picea abies* [L.] KARST) by combined gas chromatography-mass spectrometry. *J. Chromatogr.* **247**: 133~140.
- ODÉN, P. C., SCHWENEN, L. and GRAEBE, J. E. (1987) Identification of gibberellins in Norway spruce (*Picea abies*). *Plant*

Physiol. 84: 516~519.

OGIYAMA, K. and PHARIS, P. R. (1980) Endogenous plant growth regulation substance in foliage of *Cryptomeria japonica* D. DON. Mokuzai Gakkaishi 26: 823~827.

PHARIS, R. P. and KING, R. W. (1985) Gibberellins and reproductive development in seed plants. Ann. Rev. Plant Physiol. 36: 517~568.

YAMAGUCHI, I., NAKAGAWA, R., KUROGOCHI, S., MUROFUSHI, N., TAKAHASHI, N. and WEILER, E. W. (1987) Radioimmunoassay of gibberellins A₅ and A₂₀. Plant Cell Physiol. 28: 815~824.

YAMAGUCHI, I., NAKAZAWA, R., NAKAGAWA, R., SUZUKI, Y., KUROGOCHI, S., MUROFUSHI, N., TAKAHASHI, N. and WEILER, E. W. (1990) Identification and semiquantification of gibberellins from the pollen and anther of *Zea mays* by immunoassay and GC/MS. Plant Cell Physiol. 31: 1063~1069.

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