

ブロノポールを用いたアユ卵の水カビ病防除

誌名	水産増殖
ISSN	03714217
著者名	大野,平祐 畑井,喜司雄 相川,英明 原,日出夫
発行元	水産増殖談話会
巻/号	56巻1号
掲載ページ	p. 9-12
発行年月	2008年3月

農林水産省 農林水産技術会議事務局筑波産学連携支援センター
Tsukuba Business-Academia Cooperation Support Center, Agriculture, Forestry and Fisheries Research Council
Secretariat



The Use of Bronopol to Control Fungal Infections in Ayu Eggs

Heisuke OONO¹, Kishio HATAI^{1,*}, Hideaki AIKAWA² and Hideo HARA²

Abstract: We evaluated bronopol as a practical alternative anti-fungal agent to malachite green for use in hatcheries. Repeated daily exposure, from just after fertilization to the stage showing eye development, to 50 ppm and 100 ppm bronopol for 30 min showed an efficacy compared to the 0 ppm control for the inhibition of fungal infections in ayu *Plecoglossus altivelis* eggs. The 100 ppm bronopol treatment groups showed control of fungal infection up to the stage of eye development in eggs. Significant differences in terms of the number of hatched fish were seen between the 0 ppm control and treatments at 50 and 100 ppm bronopol.

Key words: Ayu eggs; Fungal control; Bronopol; Fungal infection

In Japan, fungal infections of coldwater fish and eggs have usually been controlled with malachite green. However, malachite green has never been permitted in fisheries medicine because of its teratogenic properties (Meyer and Jorgenson 1983) and the fact that the effects of its residues are still unknown (Meinertz et al. 1995). In 2005, the Pharmaceutical Affairs Law prohibited the use of malachite green in aquaculture (Miura et al. 2005). Thus, the fish culturing industry needs an alternative antifungal agent that is a safe and effective replacement for malachite green.

In Europe, bronopol has been used to control fungal infections in eggs of salmonids (Branson 2002). Bronopol (2-bromo-2-nitropropane-1, 3-diol) is water soluble, odorless and biocidal, and is widely used as a preservative in medical and pharmaceutical products such as cosmetics and shampoos (Bryce et al. 1978; Kumanova et al. 1989; Toler 1985). Bronopol is a thiol-containing dehydrogenase enzyme inhibitor that is thought to cause cell membranes to leak, consequently destroying cells (Branson 2002). This study examined the efficacy of bronopol in controlling fungal infections in ayu eggs.

Materials and Methods

Ayu eggs were taken from a single 1-year-old female and inseminated. A few minutes after insemination, approximately 100 fertilized ayu eggs were randomly attached onto 15 glass slides using a feather from a water bird. Glass slides with attached fertilized eggs were then held in metal baskets. This trial used a commercially available product, which contained 50% w/v bronopol with an inert carrier (Pyceze; Novartis Animal Vaccines Limited, Essex, United Kingdom). The eggs were treated with 50 ppm and 100 ppm bronopol for 30 min. Treatments were repeated everyday for 5 days, from fertilization until eye development was observed in the eggs. During treatment, the eggs were transferred to 2 l beakers with no aeration. Control groups, without treatment, were also prepared and transferred to beakers. After treatment, all groups were held in a 550 l tank in running water flowing at a rate of approximately 20 ml. No further treatment was performed after the eggs showed eye development. Numbers of eggs infected with fungi, dead eggs and eggs

Received July 28, 2007; Accepted November 24, 2007.

¹Division of Fish Diseases, Nippon Veterinary and Life Science University, Musashino, Tokyo 180-8602, Japan.

²Kanagawa Prefectural Fisheries Technology Center Freshwater Fisheries Experiment Station, Sagami-hara, Kanagawa 229-1135, Japan.

*Corresponding author: Tel: +81-422-31-4151, Fax: +81-422-31-6796. E-mail: hatai@nvlu.ac.jp

with eye development were counted 5 days after fertilization and numbers of eggs infected with fungi, dead eggs, hatched fish and deformities were counted 9 days after fertilization. This study was performed at the Kanagawa Prefectural Fisheries Technology Center Freshwater Fisheries Experiment Station. The source water for this hatchery was well water kept at a temperature of 20–22°C throughout the test period.

Results

The number of dead eggs with fungal infection was 22 at 0 ppm, 12 at 50 ppm bronopol and 0 at 100 ppm bronopol. The number of dead eggs without fungal infection was three at 0 ppm, 11 at 50 ppm bronopol and 25 at 100 ppm bronopol. Repeated exposure from just after fertilization up to the stage of eye development in eggs with 50 and 100 ppm bronopol showed an efficacy compared to 0 ppm for the inhibition of fungal infection (Chi-square test, $p < 0.05$). The 100 ppm bronopol displayed control of fungal infection up to the stage of eye development. The number of eggs with eyed in each group was almost same (Table 1).

Both the 50 and 100 ppm bronopol affected fungal infection from the stage of eye develop-

ment in eggs until hatching. The number of dead eggs with fungal infection was 47 at 0 ppm, 18 at 50 ppm bronopol and 12 at 100 ppm bronopol. The number of dead eggs without fungal infection was one at 0 ppm and 50 ppm bronopol, and three at 100 ppm bronopol. The number of hatched fish was 227 at 0 ppm, 271 at 50 ppm bronopol and 296 at 100 ppm bronopol. Significant differences were observed in terms of the number of hatched fish between 0 ppm and the 50 and 100 ppm bronopol (Chi-square test, $p < 0.05$; Table 2).

Discussion

The present study showed that treatment with 100 ppm bronopol for 30 min was effective in controlling fungal infections. Deformities were observed not only in groups treated with 50 ppm and 100 ppm bronopol, but also in those from the untreated group. It appeared that repeated exposure everyday, from just after fertilization to the stage of eye development in eggs, to 50 ppm and 100 ppm bronopol for 30 min, was effective in controlling fungal infections. This result was similar to the efficacy of bronopol to control fungal infections in rainbow trout *Oncorhynchus mykiss* eggs (Oono et al. 2007). The use of a fungicidal agent in hatcher-

Table 1. Control of fungal infection by bronopol from fertilization to the stage of eye development in ayu eggs ^{a)}

Concentration	Number of fertilized eggs	Dead eggs		Eggs with eyes	Unknown eggs
		With fungal infection	Without fungal infection		
0 ppm 1	58	6	0	50	2
0 ppm 2	66	3	0	63	0
0 ppm 3	65	5	0	57	3
0 ppm 4	74	3	1	69	1
0 ppm 5	70	5	2	59	4
50 ppm 1	79	0 ^{b)}	6	72	1
50 ppm 2	70	3 ^{b)}	1	65	1
50 ppm 3	58	3 ^{b)}	0	55	0
50 ppm 4	70	4 ^{b)}	3	62	1
50 ppm 5	59	2 ^{b)}	1	56	0
100 ppm 1	67	0 ^{b)}	5	60	2
100 ppm 2	87	0 ^{b)}	7	74	6
100 ppm 3	74	0 ^{b)}	3	70	1
100 ppm 4	74	0 ^{b)}	4	69	1
100 ppm 5	70	0 ^{b)}	6	64	0

Experiments were performed from September 22–27, 2004.

^{a)} Treatments were performed for 5 days from fertilization to the stage of eye development in eggs.

^{b)} Significantly different from the value for 0 ppm ($p < 0.05$).

Table 2. Control of fungal infection by bronopol from the stage of eye development to hatching in ayu eggs^{c)}

Concentration	Number of eggs with eyes	Dead eggs		Hatched fish	Deformed fish	Unknown eggs
		With fungal infection	Without fungal infection			
0 ppm 1	50	13	0	32	0	5
0 ppm 2	63	10	1	48	0	4
0 ppm 3	57	5	0	49	1	2
0 ppm 4	69	15	0	50	0	4
0 ppm 5	59	4	0	48	0	7
50 ppm 1	72	5	0	60 ^{d)}	0	7
50 ppm 2	65	3	0	58 ^{d)}	0	4
50 ppm 3	55	3	1	49 ^{d)}	0	2
50 ppm 4	62	3	0	56 ^{d)}	1	2
50 ppm 5	56	4	0	48 ^{d)}	0	4
100 ppm 1	60	5	0	48 ^{d)}	0	7
100 ppm 2	74	0	0	70 ^{d)}	1	3
100 ppm 3	70	4	0	60 ^{d)}	0	6
100 ppm 4	69	3	0	61 ^{d)}	0	5
100 ppm 5	64	0	3	57 ^{d)}	0	4

Experiments were performed from September 27-October 1, 2004.

^{c)} No treatments were performed for the 4 days from the stage of eye development to hatching.

^{d)} Significantly different from the value for 0 ppm ($p < 0.05$).

ies is usually stopped at the eye development stage of eggs. However, as it appears that fungal infections may continue from that stage to the hatched fish stage, the routine screening of dead eggs or the continuation of bronopol treatment after the eye development stage would be practical to increase hatching and control fungal infections.

References

- Branson, E. (2002) Efficacy of bronopol against infection of rainbow trout (*Oncorhynchus mikiss*) with the fungus *Saprolegnia* species. *Vet. Rec.*, **151**, 539-541.
- Bryce, D. M., B. Croshaw, J. E. Hall, V. R. Holland and B. Lessel (1978) The activity and safety of the antimicrobial agent bronopol (2-bromo-2-nitropropan-1, 3-diol). *J. Soc. Cosmet. Chems.*, **29**, 3-24.
- Hong-kyu, M., K. Hatai and B. Suk (1994) Some effects of chitosan on fish-pathogenic Oomycete, *Saprolegnia parasitica*. *Fish Pathol.*, **29**, 73-77.
- Hussein, M. M. A., S. Wada, K. Hatai and A. Yamamoto (2000) Antimycotic activities of eugenol against selected water molds. *J. Aquat. Animal Health.*, **12**, 224-229.
- Hussein, M. M. A., A. Moustafa, E. L. Feki, K. Hatai and A. Yamamoto (2002) Inhibitory effects of thymoquinone from *Nigella sativa* on *Saprolegnia* in fish. *Biocontrol Sci.*, **7**, 31-35.
- Kitancharoen, N., A. Yamamoto and K. Hatai (1997) Fungicidal effect of hydrogen peroxide on fungal infection of rainbow trout eggs. *Mycoscience*, **38**, 375-378.
- Kitancharoen, N., A. Ono, A. Yamamoto and K. Hatai (1997) The fungistatic effect of NaCl on rainbow trout egg saprolegniasis. *Fish Pathol.*, **32**, 159-162.
- Kitancharoen, N., A. Yamamoto and K. Hatai (1998) Effect of sodium chloride, hydrogen peroxide and malachite green on fungal infection of rainbow trout eggs. *Biocontrol Sci.*, **3**, 113-115.
- Kumanova, R., M. Vassileva, S. Dobрева, S. Manova and L. Kупenov (1989) Evaluating bronopol. *Manuf. Chem.*, **60**, 36-37.
- Meinertz, J. R., G. R. Stehley, W. H. Gingerich and J. L. Allen (1995) Residues of [¹⁴C]-malachite green in eggs and fry of rainbow trout, *Oncorhynchus mikiss* (Walbaum), after treatment eggs. *J. Fish Dis.*, **18**, 239-247.
- Meyer, F. P. and T. A. Jorgenson (1983) Teratological and other effects of malachite green on the development of rainbow trout and rabbit. *Tran. Amer. Fish. Soci.*, **112**, 814-824.
- Miura, M., H. Oono, N. Tuchida, K. Hatai and T. Kiryu (2005) Control of water mold infection in rainbow trout eggs by using copper fiber (in Japanese). *Fish Pathol.*, **40**, 81-86.
- Mori, T., H. Hirose, H. Chutima and K. Hatai (2002) Antifungal activities of plant extract against some aquatic fungi. *Biocontrol Sci.*, **7**, 187-191.
- Oono, H., K. Hatai, M. Miura, N. Tuchida and T. Kiryu (2007) The use of bronopol to control fungal infection in rainbow trout eggs. *Biocontrol Sci.*, **12**, 55-57.
- Pottinger, T. G. and J. G. Day (1999) A *Saprolegnia parasitica* challenge system for rainbow trout: assessment of pyceze as an anti-fungal agent for both fish and ova. *Dis. Aquat. Org.*, **36**, 129-141.
- Toler, J. C. (1985) Preservative stability and preservative systems. *Int. J. Cosmet. Sci.*, **7**, 157-164.

Yamamoto, A., S. Toyomura, M. Saneyoshi and K. Hatai (2001) Control of fungal infection of salmonid eggs by hydrogen peroxide (in Japanese). *Fish Pathol.*, **36**, 241-246.

Yuasa, K. and K. Hatai (1995) Drug sensitivity of some pathogenic water moulds isolates from freshwater fishes (in Japanese). *J. Antibact. Antifung. Agents.*, **23**, 213-219.

ブロンポールを用いたアユ卵の水カビ病防除

大野平祐・畑井喜司雄・相川英明・原 日出夫

サケ科魚類卵の水カビ病防除剤として日本で使用されてきたマラカイトグリーンは2005年で使用禁止となった。今回欧州でその代替薬として認可されたブロンポール50%含有製剤の水カビ病防除効果と毒性を検討した。試験はアユ *Plecoglossus altivelis* 受精卵を受精翌日から発眼までの5日間毎日、成分濃度で50, 100 ppm に30分間浸漬した。対照区と比較した結果、薬浴区は水カビ病の発生を阻止した。また薬浴区で孵化尾数の有意差が認められた。